

## TITLE

Periostin suppression induces decorin secretion leading to reduced breast cancer cell motility and invasion.

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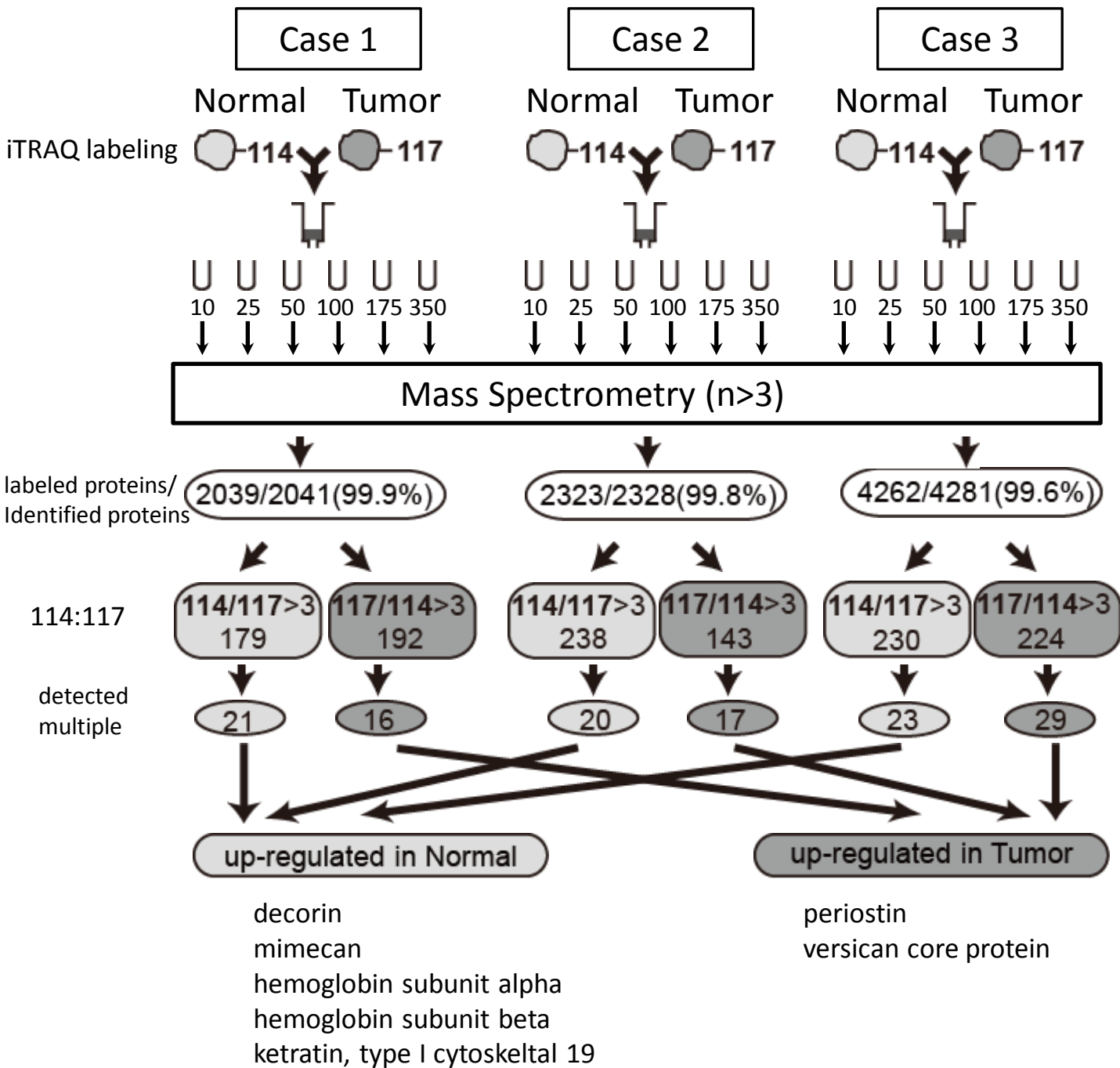
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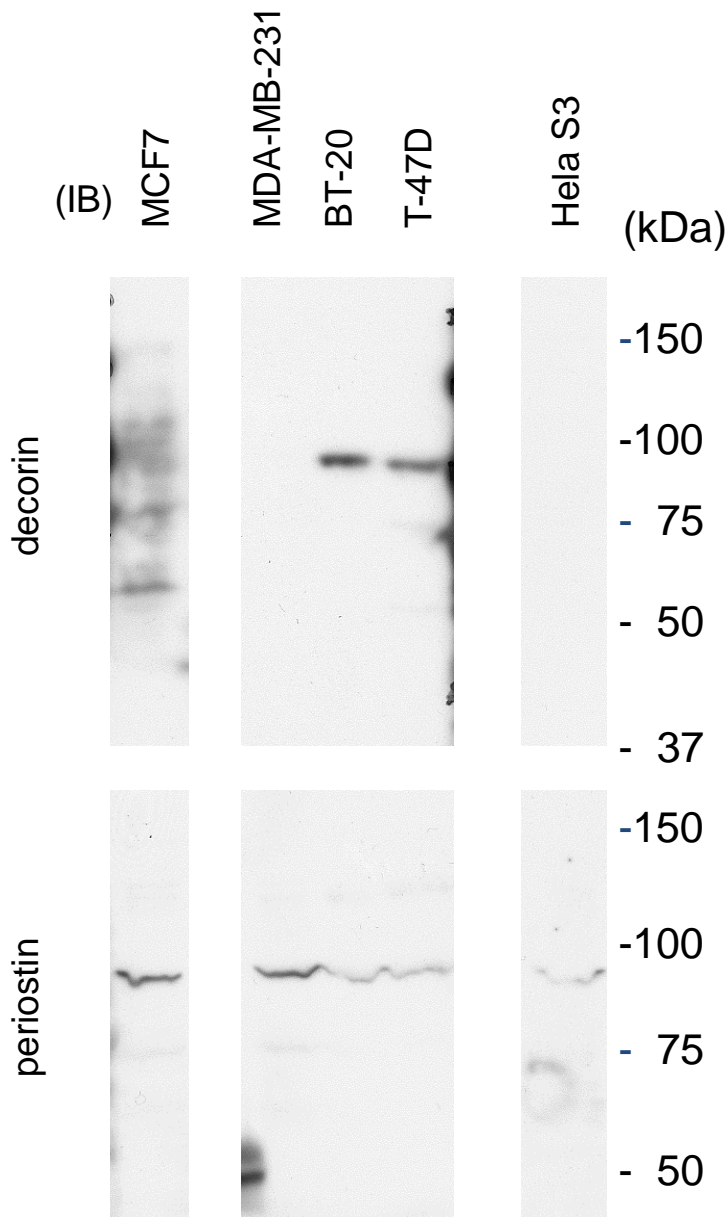
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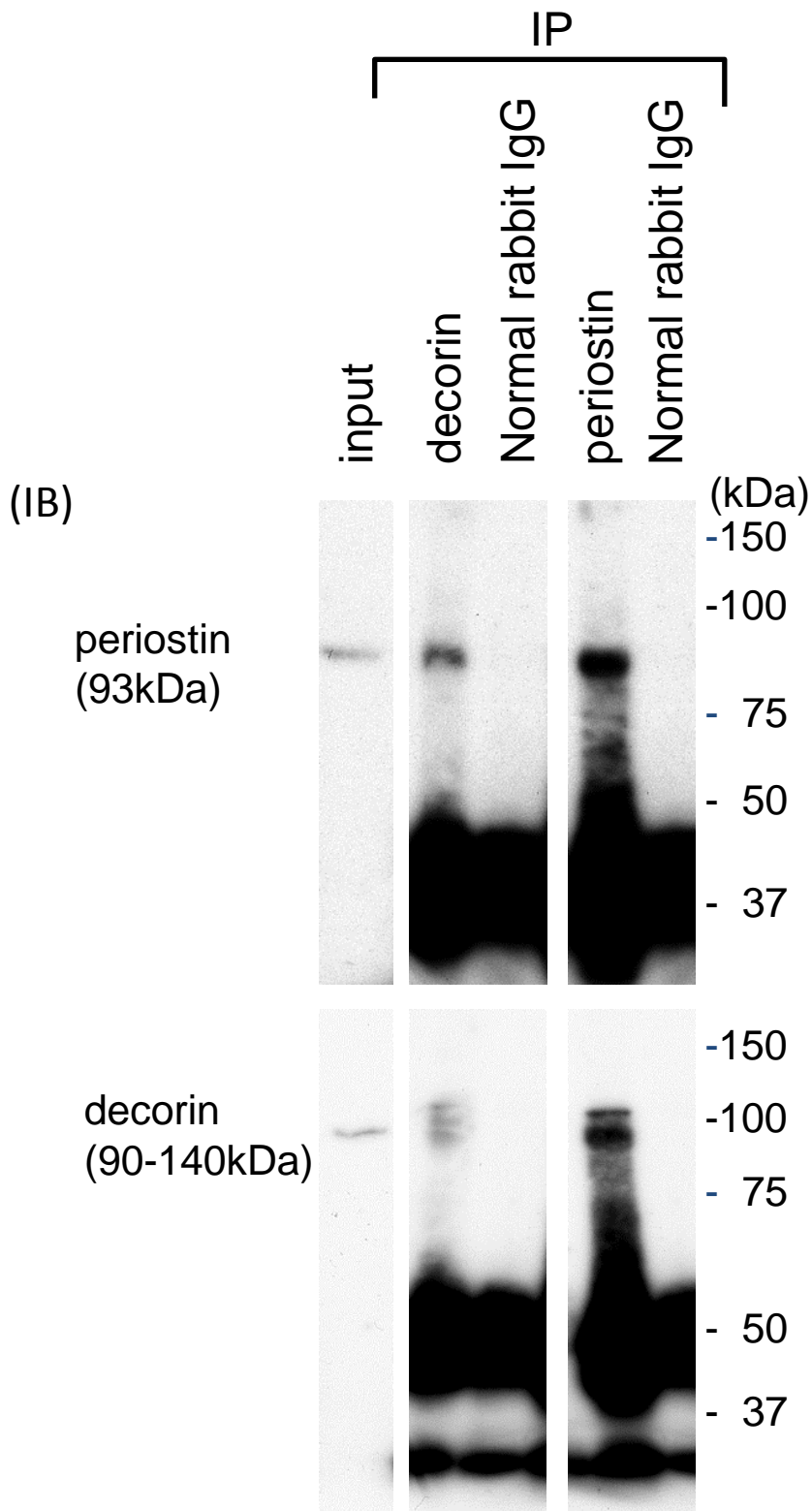
Supplementary Figure S1



**Supplementary Figure S1.** Decorin is upregulated in normal tissues relative to tumor tissues, whereas periostin is upregulated in tumor tissues. Normal and tumor tissues from three phyllodes tumor patients were depleted of albumin and subjected to trypsin digestion. Tryptic peptides were labeled using the iTRAQ reagents. Normal tissue lysates were tagged with isobaric tags with  $m/z$  of 114, and tumor tissue lysates were tagged with isobaric tags with  $m/z$  of 117. The tagged lysates were mixed and separated by chromatography on a  $C_{18}$  capillary column using elution buffer containing KCl (10, 25, 50, 100, 175, and 350 mM). The six fractions were analyzed by mass spectrometry in at least three trials.



**Supplementary Figure S2.** The levels of expression of decorin and periostin in the breast cancer cell lines. Cell lysates from cancer cell lines MCF7, MDA-MB-231, BT-20, T-47D and HeLaS3 were immunoblotted using anti-decorin and anti-periostin antibodies. These cropped blots are used in the main figure(Figure 3a).

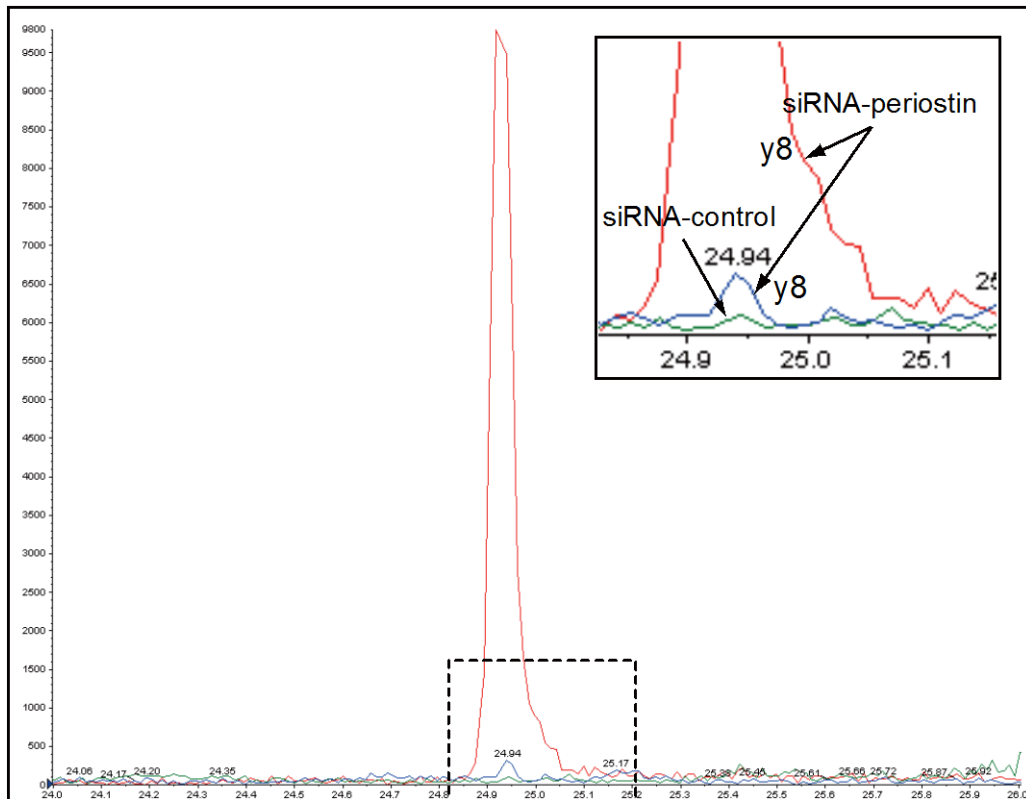


**Supplementary Figure S3.** Plasma membrane translocation of decorin upon knockdown of periostin in BT-20 cells. Anti-periostin or anti-decorin immunoprecipitates from BT-20 cell lysates were subjected to SDS-PAGE, followed by immunoblot analysis with anti-decorin or anti-periostin, respectively. Input lysate was used as a positive control, and normal rabbit IgG was used as a negative control. Tissue lysates were immunoblotted with anti-decorin and anti-periostin antibodies. N, normal tissue; T, tumor tissue. These cropped blots are used in the main figure (Figure 3b).

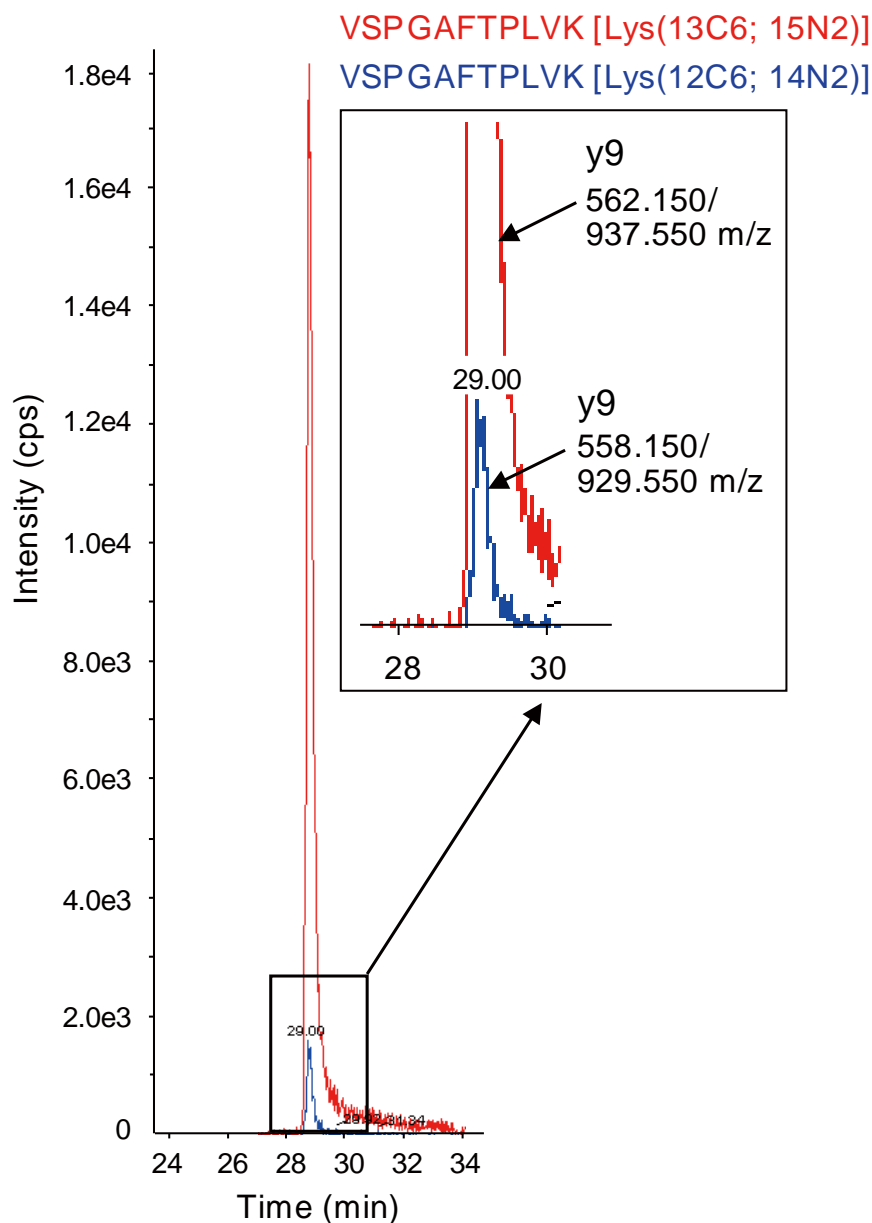
DLPPDTLLDLQNNK [Lys (12C6, 14N2); 848.670 m/z (Q1), 957.480 m/z (Q3)]

DLPPDTLLDLQNNK [Lys (13C6, 15N2); 852.670 m/z (Q1), 965.480 m/z (Q3)]

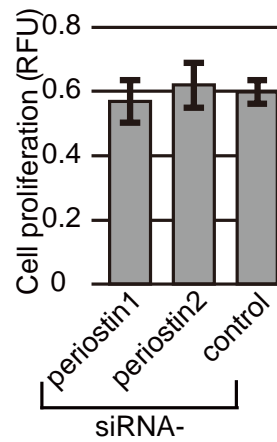
DLPPDTLLDLQNNK [Lys (12C6, 14N2); 848.670 m/z (Q1), 957.480 m/z (Q3)]



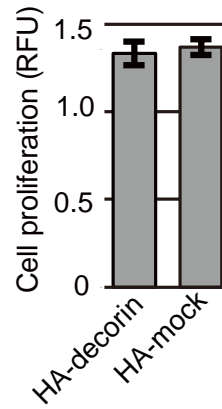
**Supplementary Figure S4.** MRM chromatograms for DLPPDTLLDLQNNK fragments and their standard (AQUA: red line) analogues. MRM chromatograms for VSPGAFTPLVK was shown in Figure 4a. The peptides in culture medium from siRNA-periostin-treated cells (blue line) or siRNA-control-treated cells (green line) were analyzed using the MRM method. The doubly charged precursor mass was chosen as the Q1 mass, and the y8 fragment ion was chosen as the Q3 mass. Insets contain magnified views.



**Supplementary Figure S5.** MRM chromatograms for VSPGAFTPLVK fragments and their standard (AQUA) analogues. Two MRM transitions for the endogenous (blue line) and standard (red line) peptides were monitored (Figure 4b). For VSPGAFTPLVK, the doubly charged precursor mass was chosen as the Q1 mass, and the y9 fragment ions were chosen as Q3 mass. Insets contain magnified views. MRM-triggered MS/MS product ion spectra obtained by nanoflow LC/MS/MS, comparing normal tissue digested with trypsin (Figure 4c; upper) with decorin secreted from decorin-overexpressing MDA-MB-231 cells (Figure 4c; lower). The spectrum of the peptide clearly shows y-ion fragments.



**Supplementary Figure S6.** Proliferation of BT-20 cells treated with siRNA-periostin or siRNA-control. Cell number was measured using water-soluble tetrazolium-1 (WST-1) assay. Relative fluorescence units (RFU) indicate the relative amount of proliferation. Column graphs show the means  $\pm$  SEM of results from six samples.



**Supplementary Figure S7.** Proliferation of MDA-MB-231 transiently expressing HA-decorin or HA-mock in MDA-MB-231 cells was measured using the WST-1 assay. Column graphs show means  $\pm$  SEM of results from seven samples.





# Supplementary Table S2

IP:decorin

		Unused	% Cov	Name	Peptides(95%)	
1	case1	Normal	9.67	48.5	Decorin	4
		Tumor	4.06	48.2	Decorin	1
	case2	Normal	14.06	53.8	Decorin	7
		Tumor	4.42	59.3	Decorin	2
	case3	Normal	18.61	61	Decorin	10
Tumor		14.1	50.1	Decorin	7	
2	case1	Normal	3.65	28.6	Periostin	1
			2.55	53.2	Decorin	1
		Tumor	11.45	43.4	Periostin	5
			7.19	28.1	Decorin	3
	case2	Normal	20.75	46.5	Decorin	13
		Tumor	4.06	32	Decorin	2
	case3	Normal	11.74	56.8	Decorin	5
			2.06	39.1	Dermcidin	1
Tumor		1.7	13.6	Hornerin	1	
3	case1	Normal	2	35.1	60 kDa heat shock protein, mitochondrial	1
		Tumor	10.39	30.1	Alpha-1-antitrypsin	6
	case2	Tumor	2.21	22	Alpha-1-antitrypsin	1
			2	22.5	60 kDa heat shock protein, mitochondrial	2
	case3	Tumor	2.74	32.3	60 kDa heat shock protein, mitochondrial	1
			2.46	31.3	Alpha-1-antitrypsin	1
4	case1	Normal	11.91	43.2	Actin, cytoplasmic 1	6
			8.26	65.2	Vimentin	3
			0.35	23.9	Beta-actin-like protein 2	1
	case2	Normal	8.98	40	Actin, cytoplasmic 1	4
			15.01	50.1	Actin, cytoplasmic 2	5
			3.34	40.3	Vimentin	2
	case3	Tumor	2.53	36.2	Decorin	1
			14.93	49.1	Actin, cytoplasmic 2	8
			2	49.1	Actin, cytoplasmic 1	8
			11.1	42.9	Actin, cytoplasmic 1	4
	case3	Tumor	16.13	56.3	Actin, cytoplasmic 1	8
			0.64	56	Actin, cytoplasmic 2	7
			0.46	38.8	Beta-actin-like protein 2	2
0.18			54.1	Actin, aortic smooth muscle	3	

IP:periostin

		Unused	% Cov	Name	Peptides(95%)	
5	case1	Normal	1.3	11.3	Voltage-dependent L-type calcium channel subunit alpha-1C	1
		Tumor	2	12.8	Alpha-2-macroglobulin	1
6	case1	Tumor	4.26	31.8	Decorin	2
	case2	Normal	28.51	73.8	Decorin	16
		Tumor	3.54	35.4	Decorin	2
	case3	Normal	2.42	27.6	Decorin	1
		Tumor	13.7	39.6	Decorin	5
7	case1	Normal	17.74	46.2	Decorin	8
			14.54	42.1	Periostin	4
		Tumor	25.97	55.1	Periostin	10
			4.15	34.3	Decorin	2
	case2	Normal	41.92	53.2	Decorin	23
			3.55	22.4	Periostin	1
		Tumor	14.93	37.3	Periostin	8
			3.79	36.2	Decorin	2
	case3	Tumor	2.38	43.6	Dermcidin	1
			3.56	33.7	Decorin	1
7.84			37.9	Decorin	3	
5.42			33.1	Periostin	2	
8	case1	Normal	1.47	18.6	Lumican	1
			1.36	30	Putative V-set and immunoglobulin domain-containing-like protein ENSP00000303034	1
	case2	Tumor	10.72	45.7	Alpha-1-antitrypsin	3
			2.07	28.7	Alpha-1-antitrypsin	1
case3	Tumor	2	14.8	60 kDa heat shock protein, mitochondrial	1	
9	case1	Normal	1.3	12.2	Nuclear pore membrane glycoprotein 210	1

**Supplementary Table S2.** List of decorin- or periostin-binding target proteins from Figure 3e and f.

## Supplementary Table S3

cell	medium
BT-20	Eagle's Minimum Essential Medium with L-Glutamine (ATCC, Manassas, VA, USA) supplemented with 10% fetal bovine serum
T-47D	RPMI-1640 with High Glucose, L-glutamine, and HEPES (ATCC) supplemented with 10% fetal bovine serum and insulin
MDA-MB-231	DMEM/F12 (1:1) with L-Glutamine (Life Technologies, Grand Island, NY, USA) supplemented with 10% fetal bovine serum and HEPES
MCF7	Eagle's Minimum Essential Medium with 10% fetal bovine serum and 0.01 mg/ml human recombinant insulin
HeLa S3	DMEM with 10% fetal bovine serum, NaHCO <sub>3</sub> , and L-glutamine

**Supplementary Table S3.** List of the mediums for each cell line.

## Supplementary Table S4

Application	antibody	species	source
Immunoblotting	decorin	mouse	Santa Cruz Biotechnology, Dallas, TX, USA
	periostin	rabbit	Abcam, Cambridge, UK
	E-cadherin	mouse	Takara, Otsu, Japan
	$\beta$ -actin	mouse	Sigma-Aldrich, St. LouisMO, USA
Immunoprecipitation	decorin	rabbit	Bioss, Woburn, MA, USA
	periostin	rabbit	Santa Cruz Biotechnology, Dallas, TX, USA
Immunohistochemical staining	decorin	mouse	Abcam, Cambridge, UK
	periostin	rabbit	Abcam, Cambridge, UK
Immunofluorescence	decorin	mouse	Abcam, Cambridge, UK
	periostin	rabbit	GeneTex, Irvine, CA, USA

**Supplementary Table S4.** List of antibodies.

## Supplementary Method

### Immunoblotting analysis

Tissue and cell lysates were subjected to SDS-PAGE and transferred electrophoretically onto Immobilon-P transfer membrane (EMD Millipore, Billerica, MA, USA). The membrane was incubated in SuperBlock Blocking Buffer (Thermo Scientific, Waltham, MA, USA) at room temperature (RT) for 1 h, stained with primary antibodies for 2 h at RT, and incubated for 1 h at RT with secondary antibodies coupled to horseradish peroxidase (ECL anti-mouse or anti-rabbit IgG; Amersham/GE Healthcare, Buckinghamshire, UK). The blots were developed using the SuperSignal enhanced chemiluminescence reagent (Pierce, Rockford, IL, USA) and exposed to Kodak X-OMAT film. All antibodies are listed in Supplementary Table S4. As needed, protein bands were quantitated using the ImageJ software (version 1.46r, National Institutes of Health, Bethesda, MD, USA).

### Immunoprecipitation

Tissue and cell lysates were immunoprecipitated for 30 min at 4° C with 5 µg antibody, followed by incubation for 30 min at 4° C with Dynabeads–Protein G (Invitrogen, Carlsbad, CA, USA). Immune complexes were precipitated and washed. Immunoprecipitated proteins were loaded to onto e-PAGEL precast gels (ATTO, Tokyo, Japan) for mass spectrometry or subjected to standard SDS-PAGE for immunoblotting.

### Knockdown of gene expression

siRNAs specifically targeting periostin and decorin were purchased from Dharmacon (Lafayette, CO, USA). For transfections, Lipofectamine RNAiMAX (Invitrogen) and 12 nM siRNA were mixed in 2 ml Opti-MEM (Life Technologies) and incubated for 20 minutes at RT; the mixture was then added onto 70% confluent cultures. After 72 h, RNA and protein were isolated.

### Total RNA extraction and RT-PCR

Total RNA was extracted from the lysates BT-20 using the QuickPrep Micro mRNA Purification Kit (GE Healthcare, Little Chalfont, UK). Decorin full-length cDNA was generated from RNA by RT-PCR using the SuperScript III One-Step RT-PCR System with Platinum Taq (Life Technologies) and subcloned into the pME18S-HA-C vector in frame with the appropriate N-terminal tags. Clonal cell lines were obtained by transfection of 2 µg of plasmid DNA using the Lipofectamine Plus transfection reagent (Invitrogen). Reverse transcription was performed in a reaction volume of 60 µl containing 90 ng of total RNA and 2 µl of SuperScript III One-Step RT-PCR System with Platinum Taq. The conditions for polymerase chain reaction (PCR) were as follows: 94° C for 2 min; 40 cycles of 94° C for 15 s, 60° C for 30 s, and 68° C for 90 s; and a final cycle of 68° C for 5 min. The PCR products were electrophoresed on 1% agarose gels. The PCR primers for *DCN* were 5'-CCGGAATTCATGAAGGCCACTATCATCC-3' and 5'-GCCGCTCGAGTTACTTATAGTTTCCGAG-3'.

### Immunofluorescence

Cells were seeded onto chamber slides (Matsunami, Osaka, Japan) and fixed with 3.5% formaldehyde in PBS for 10 minutes on ice. Fixed cells were permeabilized with 50%, 75%, and 95% cold ethanol on ice for 5 minutes each. Fixed cells were blocked for 30 min, incubated with primary antibody for 1 h at RT, incubated with Alexa Fluor 488- or Alexa Fluor 594-conjugated secondary antibody (Molecular Probes, Life Technologies) for 30 min at 37° C, and then preserved in Vectashield (Vector Laboratories, Burlingame, CA, USA). DNA was stained with bisbenzimidazole (Hoechst 33258, Life Technologies). The samples were examined with an Olympus Power BX51 fluorescence microscope (Olympus, Tokyo, Japan). Confocal microscopy was performed using a Leica TS SP8, and the confocal images were analyzed using the Leica SP8 LAS AF Lite software (Leica, Solms, Germany).

### Cell proliferation assay

Cells were seeded in 96-well flat-bottomed microtiter plates and cultured for 24 h, followed by incubation for 30 minutes with Cell Proliferation Reagent WST-1 (Roche). Cleavage of WST-1 to formazan by metabolically active cells was quantitated by scanning the plates at 450 nm and 650 nm (reference wavelength) in a microtiter plate reader. Viability of treated cells was normalized to that of untreated control cells.