**Experimental design**

1. **Sample size**
   
   Describe how sample size was determined.
   
   For single-candidate luciferase assays, we performed experiments in triplicates (independent transfections), as is standard in the field. For NGS experiments, we performed two replicates, as is standard in the field, except if equivalent experiments independently confirmed the results.

2. **Data exclusions**
   
   Describe any data exclusions.
   
   No data was excluded.

3. **Replication**
   
   Describe whether the experimental findings were reliably reproduced.
   
   All replication attempts were successful.

4. **Randomization**
   
   Describe how samples/organisms/participants were allocated into experimental groups.
   
   Not relevant because the samples were not grouped.

5. **Blinding**
   
   Describe whether the investigators were blinded to group allocation during data collection and/or analysis.
   
   All experiments were done in cell culture and did not involve animal or human research participants and blinding did therefore not apply.

   Note: all studies involving animals and/or human research participants must disclose whether blinding and randomization were used.

6. **Statistical parameters**
   
   For all figures and tables that use statistical methods, confirm that the following items are present in relevant figure legends (or in the Methods section if additional space is needed).

   - The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement (animals, litters, cultures, etc.)
   - A description of how samples were collected, noting whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
   - A statement indicating how many times each experiment was replicated
   - The statistical test(s) used and whether they are one- or two-sided (note: only common tests should be described solely by name; more complex techniques should be described in the Methods section)
   - A description of any assumptions or corrections, such as an adjustment for multiple comparisons
   - The test results (e.g. P values) given as exact values whenever possible and with confidence intervals noted
   - A clear description of statistics including central tendency (e.g. median, mean) and variation (e.g. standard deviation, interquartile range)
   - Clearly defined error bars

   See the web collection on statistics for biologists for further resources and guidance.
### Software

Policy information about availability of computer code

#### 7. Software

Describe the software used to analyze the data in this study.

- **Read mapping:** bowtie version 0.12.9. Coverage calculation, region intersection: bedtools 2.19.1. Peak annotation: R GenomicRanges 1.20.8. Reservoir sampling: sample 1.0.2. Gene ontology: R topGO 2.20.0. Disease ontology: GREAT 3.0.0. Motif analysis: MAST/MEME 4.8.1. Heatmaps: Java TreeView 1.6.4. RNA-seq: kallisto 0.43.0. Clustering/Heatmaps: R pheatmap 1.0.8. All other tasks were performed using custom shell and R scripts, which are available upon request.

For manuscripts utilizing custom algorithms or software that are central to the paper but not yet described in the published literature, software must be made available to editors and reviewers upon request. We strongly encourage code deposition in a community repository (e.g. GitHub). Nature Methods guidance for providing algorithms and software for publication provides further information on this topic.

### Materials and reagents

Policy information about availability of materials

#### 8. Materials availability

Indicate whether there are restrictions on availability of unique materials or if these materials are only available for distribution by a for-profit company.

- All plasmids are available from Addgene upon publication.

#### 9. Antibodies

Describe the antibodies used and how they were validated for use in the system under study (i.e. assay and species).

- No antibodies were used.

#### 10. Eukaryotic cell lines

- **a.** State the source of each eukaryotic cell line used.
  - HeLa-S3, HCT-116, SK-N-SH cells were purchased from ATCC (cat# CCL-2.2, CCL-247, CCL-243, HTB-11) and GM12878 cells from Coriell Institute. K562 were a gift from the Zuber lab (IMP) and THP-1 and U937 cells were a gift from the Decker and Versteeg labs (MPFL).

- **b.** Describe the method of cell line authentication used.
  - As the cell lines were purchased directly from ATCC and Coriell, visual inspection was used to confirm the morphologies of the cell lines (compared to pictures provided by the vendors). K562 cells were confirmed by STR authentication. THP-1 and U937 cells were not further authenticated.

- **c.** Report whether the cell lines were tested for mycoplasma contamination.
  - All cell lines except for THP-1 and U937 cells were tested for mycoplasma and tested negative.

- **d.** If any of the cell lines used are listed in the database of commonly misidentified cell lines maintained by ICLAC, provide a scientific rationale for their use.
  - U937 are one of the most commonly used cell types in basic research, which is why it was important to include them in the panel of cell lines we screened for supplemental figure S2C.

### Animals and human research participants

Policy information about studies involving animals; when reporting animal research, follow the ARRIVE guidelines

#### 11. Description of research animals

Provide details on animals and/or animal-derived materials used in the study.

- No animals were used.

Policy information about studies involving human research participants

#### 12. Description of human research participants

Describe the covariate-relevant population characteristics of the human research participants.

- No human participants were involved.