Experimental design

1. Sample size
   Describe how sample size was determined.
   No statistical methods were used to predetermine sample size. During experiments, we determined that a sample size of about 10-15 was sufficient to assess whether the order of differences between conditions expected from our computational model were found.

2. Data exclusions
   Describe any data exclusions.
   No data were excluded from the analyses.

3. Replication
   Describe whether the experimental findings were reliably reproduced.
   All experimental findings were reproduced independently at least two times.

4. Randomization
   Describe how samples/organisms/participants were allocated into experimental groups.
   Cells were measured at random within each condition.

5. Blinding
   Describe whether the investigators were blinded to group allocation during data collection and/or analysis.
   N/A

Note: all studies involving animals and/or human research participants must disclose whether blinding and randomization were used.

6. Statistical parameters
   For all figures and tables that use statistical methods, confirm that the following items are present in relevant figure legends (or in the Methods section if additional space is needed).

   - The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement (animals, litters, cultures, etc.)
   - A description of how samples were collected, noting whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
   - A statement indicating how many times each experiment was replicated
   - The statistical test(s) used and whether they are one- or two-sided (note: only common tests should be described solely by name; more complex techniques should be described in the Methods section)
   - A description of any assumptions or corrections, such as an adjustment for multiple comparisons
   - The test results (e.g. P values) given as exact values whenever possible and with confidence intervals noted
   - A clear description of statistics including central tendency (e.g. median, mean) and variation (e.g. standard deviation, interquartile range)
   - Clearly defined error bars

   See the web collection on statistics for biologists for further resources and guidance.
7. Software

Describe the software used to analyze the data in this study.

Custom code in Matlab was employed to implement the computational clutch model, and to calculate traction forces.

For manuscripts utilizing custom algorithms or software that are central to the paper but not yet described in the published literature, software must be made available to editors and reviewers upon request. We strongly encourage code deposition in a community repository (e.g. GitHub). *Nature Methods guidance for providing algorithms and software for publication* provides further information on this topic.

8. Materials availability

Indicate whether there are restrictions on availability of unique materials or if these materials are only available for distribution by a for-profit company.

No unique materials were used.

9. Antibodies

Describe the antibodies used and how they were validated for use in the system under study (i.e. assay and species).

α5β1 blocking integrin blocking antibody (30μg/ml, clone JBS5 - MAB1969, Millipore)
Phospho-paxillin antibody(Cell Signaling 2541S, 1:50 dilution)
YAP antibody (clone 63.7 produced in mouse, Santa Cruz catalogue no. sc-101199, 1:200 dilution)

All antibodies are validated for the species and assay used as described in the manufacturer's web page.

10. Eukaryotic cell lines

a. State the source of each eukaryotic cell line used.

Human breast myoepithelial immortalized cell lines were described previously (supplementary refs 4,5 of the paper). Human umbilical vein endothelial cells were purchased from Lonza (CC-2517). Mouse Embryonic fibroblasts were described previously (supplementary ref. 6 of the paper). Mammary epithelial cells (MCF10A) were purchased from ATCC.

b. Describe the method of cell line authentication used.

Myoepithelial cells used throughout the manuscript were authenticated in their lab of origin through expression of Integrin β4, P-cadherin, cytokeratin 17, and desmoglein 3. Other cell lines (used only to verify the generality of our findings) were not authenticated.

c. Report whether the cell lines were tested for mycoplasma contamination.

All cell lines tested negative for mycoplasma contamination.

d. If any of the cell lines used are listed in the database of commonly misidentified cell lines maintained by ICLAC, provide a scientific rationale for their use.

No commonly misidentified cell lines were used.

11. Description of research animals

Provide details on animals and/or animal-derived materials used in the study.

No animals were used.

12. Description of human research participants

Describe the covariate-relevant population characteristics of the human research participants.

This study did not involve human research participants.