

cachexia affects survival in cancer, if progress could be made to stop tissue wasting, it would substantially alleviate the disease burden for patients. ■

**J. Matthias Löhr** is in the Department of Cancer Medicine, Karolinska University Hospital, and in the Department of Clinical Intervention and Technology, Karolinska Institutet, 141 86 Stockholm, Sweden. e-mail: matthias.lohr@ki.se

1. DiMagno, E. P. *Ann. Oncol.* **10** (Suppl. 4),

- S140–S142 (1999).  
 2. Danai, L. V. *et al. Nature* **558**, 600–604 (2018).  
 3. Tisdale, M. J. *Nature Rev. Cancer* **2**, 862–871 (2002).  
 4. Fearon, K., Arends, J. & Baracos, V. *Nature Rev. Clin. Oncol.* **10**, 90–99 (2013).  
 5. Milton, K. J. *Nutr.* **133**, 3886S–3892S (2003).  
 6. Vujasinovic, M., Valente, R., Del Chiaro, M., Permert, J. & Löhr, J. M. *Nutrients* **9**, 183 (2017).  
 7. Murphy, R. A. *et al. Lipids* **47**, 363–369 (2012).  
 8. Dev, R. *et al. Oncologist* **16**, 1637–1641 (2011).  
 9. Abe, K. *et al. Anticancer Res.* **38**, 2369–2375 (2018).  
 10. Mayers, J. R. *et al. Nature Med.* **20**, 1193–1198 (2014).  
 11. Michaelis, K. A. *et al. J. Cachexia Sarcopenia Muscle* **8**, 824–838 (2017).  
 12. Hingorani, S. R. *et al. Cancer Cell* **4**, 437–450 (2003).

13. Di Sebastiano, K. M. *et al. Br. J. Nutr.* **109**, 302–312 (2013).  
 14. Gupta, S. *et al. Clin. Gastroenterol. Hepatol.* **4**, 1366–1372 (2006).  
 15. Kalyani, R. R., Corriere, M. & Ferrucci, L. *Lancet Diabetes Endocrinol.* **2**, 819–829 (2014).  
 16. Hardt, P. D. *et al. Pancreatology* **3**, 395–402 (2003).  
 17. Wagner, E. F. & Petruzzelli, M. *Nature* **521**, 430–431 (2015).  
 18. Greco, S. H. *et al. PLoS ONE* **10**, e0132786 (2015).  
 19. Löhr, J. M., Panic, N., Vujasinovic, M. & Verbeke, C. S. *J. Intern. Med.* **283**, 446–460 (2018).

The author declares competing financial and other interests. See [go.nature.com/2sp7yeo](https://go.nature.com/2sp7yeo) for details.

This article was published online on 20 June 2018.

## In retrospect

# Twenty years of network science

The idea that everyone in the world is connected to everyone else by just six degrees of separation was explained by the ‘small-world’ network model 20 years ago. What seemed to be a niche finding turned out to have huge consequences.

ALESSANDRO VESPIGNANI

In 1998, Watts and Strogatz<sup>1</sup> introduced the ‘small-world’ model of networks, which describes the clustering and short separations of nodes found in many real-life networks. I still vividly remember the discussion I had with fellow statistical physicists at the time: the model was seen as sort of interesting, but seemed to be merely an exotic departure from the regular, lattice-like network structures we were used to. But the more the paper was assimilated by scientists from different fields, the more it became clear that it had deep implications for our understanding of dynamic behaviour and phase transitions in real-world phenomena ranging

from contagion processes to information diffusion. It soon became apparent that the paper had ushered in a new era of research that would lead to the establishment of network science as a multidisciplinary field.

Before Watts and Strogatz published their paper, the archetypical network-generation algorithms were based on construction processes such as those described by the Erdős–Rényi model<sup>2</sup>. These processes are characterized by a lack of knowledge of the principles that guide the creation of connections (edges) between nodes in networks, and make the simple assumption that pairs of nodes can be connected at random with a given connection probability. Such a process generates random networks, in which the

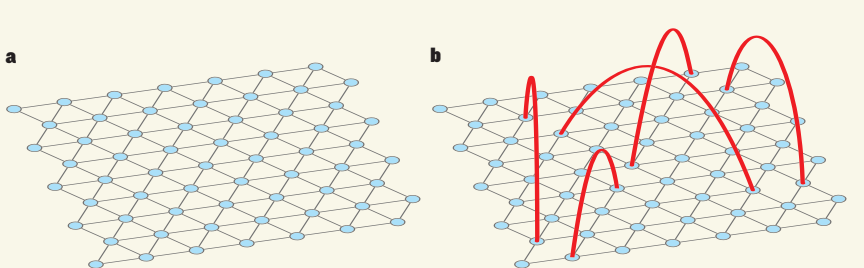
average path length between any two nodes in the network — measured as the smallest number of edges needed to connect the nodes — scales as the logarithm of the total number of nodes. In other words, randomness is sufficient to explain the small-world phenomenon popularized as ‘six degrees of separation’<sup>3,4</sup>: the idea that everyone in the world is connected to everyone else through a chain of, at most, six mutual acquaintances.

However, random construction fell short of capturing the local cliquishness of nodes observed in real-world networks. Cliquishness is measured quantitatively by the clustering coefficient of a node, which is defined as the ratio of the number of links between a node’s neighbours and the maximum number of such links. In real-world networks, node clustering is clearly exemplified by the axiom ‘the friends of my friends are my friends’: the probability of three people being friends with each other in a social network, for example, is generally much higher than would be predicted by a model network constructed using the simple, stochastic process.

To overcome the dichotomy between randomness and cliquishness, Watts and Strogatz proposed a model whose starting point is a regular network that has a large clustering coefficient. Stochasticity is then introduced by allowing links to be rewired at random between nodes, with a fixed probability of rewiring ( $p$ ) for all links. By tuning  $p$ , the model effectively interpolates between a regular lattice ( $p \rightarrow 0$ ) and a completely random network ( $p \rightarrow 1$ ).

At very small  $p$  values, the resulting network is a regular lattice and therefore has a high clustering coefficient. However, even at small  $p$ , short cuts appear between distant nodes in the lattice, dramatically reducing the average shortest path length (Fig. 1). Watts and Strogatz showed that, depending on the number of nodes<sup>5</sup>, it is possible to find networks that have a large clustering coefficient and short average distances between nodes for a broad range of  $p$  values, thus reconciling the small-world phenomenon with network cliquishness.

Watts and Strogatz’s model was initially regarded simply as the explanation for six degrees of separation. But possibly its most important impact was to pave the way for



**Figure 1 | The small-world network model.** In 1998, Watts and Strogatz<sup>1</sup> described a model that helps to explain the structures of networks in the real world. **a**, They started with a regular network, depicted here as nodes connected in a triangular lattice in which each node is connected to six other nodes. **b**, They then allowed links between nodes to be rewired at random, with a fixed probability of rewiring for all links. As the probability increases, an increasing number of short cuts (red lines) connect distant nodes in the network. This generates the small-world effect: all nodes in the network can be connected by passing along a small number of links between nodes, but neighbouring nodes are connected to one another, forming clustered cliques. (Adapted from Samay/Vespignani.)

studies of the effect of network structure on a wide range of dynamic phenomena. Another paper was also pivotal: in 1999, Barabási and Albert proposed the 'preferential-attachment' network model<sup>6</sup>, which highlighted that the probability distribution describing the number of connections that form between nodes in real-world networks is often characterized by 'heavy-tailed' distributions, instead of the Poisson distribution predicted by random networks. The broad spectrum of emergent behaviour and phase transitions encapsulated in networks that have clustered connectedness (as in Watts and Strogatz's model) and heterogeneous connectedness (as in the preferential-attachment model) attracted the attention of scientists from many fields.

A string of discoveries followed, highlighting how the complex structure of such networks underpins real-world systems, with implications for network robustness, the spreading of epidemics, information flow and the synchronization of collective behaviour across networks<sup>7,8</sup>. For example, the small-world connectivity pattern proved to be the key to understanding the structure of the World Wide Web<sup>9</sup> and how anatomical and functional areas of the brain communicate with each other<sup>10</sup>. Other structural properties of networks came under the microscope soon after<sup>11–13</sup>, such as modularity and the concept of structural motifs, all of which helped scientists to characterize and understand the architecture of living and artificial systems, from subcellular networks to ecosystems and the Internet.

The current generation of network research cross-fertilizes areas that benefit from unprecedented computing power, big data sets and new computational modelling techniques, and thus provides a bridge between the dynamics of individual nodes and the emergent properties of macroscopic networks. But the immediacy and the simplicity of the small-world and preferential-attachment models still underpin our understanding of network topology. Indeed, the relevance of these models to different areas of science laid the foundation of the multidisciplinary field now known as network science.

Integrating knowledge and methodologies from fields as disparate as the social sciences, physics, biology, computer science and applied mathematics was not easy. It took several years to find common ground, agree on definitions and reconcile and appreciate the different approaches that each field had adopted to study networks. This is still a work in progress, presenting all the difficulties and traps inherent in interdisciplinary work. However, in the past 20 years a vibrant network-science community has emerged, with its own prestigious journals, research institutes and conferences attended by thousands of scientists.

By the 20th anniversary of the paper, more than 18,000 papers have cited the model, which is now considered to be one of the benchmark network topologies. Watts and Strogatz closed

their paper by saying: "We hope that our work will stimulate further studies of small-world networks." Perhaps no statement has ever been more prophetic. ■

**Alessandro Vespignani** is in the Network Science Institute and the Laboratory for the Modeling of Biological and Sociotechnical Systems, Northeastern University, Boston, Massachusetts 02115, USA.  
e-mail: a.vespignani@northeastern.edu

1. Watts, D. J. & Strogatz, S. H. *Nature* **393**, 440–442 (1998).
2. Erdős, P. & Rényi, A. *Publ. Math.* **6**, 290–297 (1959).
3. Milgram, S. *Psychol. Today* **1**, 61–67 (1967).

## STRUCTURAL BIOLOGY

# A complex story of receptor signalling

**G-protein-coupled receptors activate different G-protein types to trigger divergent signalling pathways. Four structures of receptor–G-protein complexes shed light on this selectivity. SEE ARTICLES P.547, P.553 & P.559 & LETTER P.620**

MICHAEL J. CAPPER & DANIEL WACKER

About one-third of all drugs, including opioid painkillers, antihistamines and many antipsychotics, target members of a family of proteins called G-protein-coupled receptors (GPCRs)<sup>1</sup>. This reflects the fact that GPCRs are important in almost all aspects of human physiology, and suggests that many more of them will be promising drug targets for numerous diseases. GPCRs span the cell membrane and convert myriad extracellular signals, including neurotransmitter molecules, hormones, and even light, into a cellular response by activating cellular G proteins and other transducer proteins. Four papers<sup>2–5</sup> in this issue help to unravel the mystery of how GPCRs selectively activate a particular group of G proteins known as G<sub>i/o</sub>, and provide clues that might aid the design of improved GPCR-targeting drugs.

Although more than 800 GPCRs are encoded in the human genome, they couple to only a small number of intracellular signal transducers, including 16 Gα proteins<sup>6</sup>. The latter proteins assemble with Gβ and Gγ proteins to form heterotrimeric G proteins. The G-protein complex disassembles on activation by GPCRs, whereupon the various subunits activate different signalling pathways. For instance, stimulatory Gα proteins (known as G<sub>s</sub>) increase cellular levels of cyclic AMP molecules, which regulate various cellular processes. Structures of G<sub>s</sub>-bound GPCRs have been reported<sup>7,8</sup> that have begun to elucidate the general activation mechanism of Gα proteins, and of G<sub>s</sub> in

4. Guare, J. *Six Degrees of Separation* (Vintage, 1990).
5. Barthélémy, M. & Amaral, L. A. N. *Phys. Rev. Lett.* **82**, 3180–3183 (1999).
6. Barabási, A.-L. & Albert, R. *Science* **286**, 509–512 (1999).
7. Pastor-Satorras, R., Castellano, C., Van Mieghem, P. & Vespignani, A. *Rev. Mod. Phys.* **87**, 925–979 (2015).
8. Arenas, A., Díaz-Guilera, A., Kurths, J., Moreno, Y. & Zhou, C. *Phys. Rep.* **469**, 93–153 (2008).
9. Albert, R., Jeong, H. & Barabási, A.-L. *Nature* **401**, 130–131 (1999).
10. Sporns, O., Chialvo, D. R., Kaiser, M. & Hilgetag, C. C. *Trends Cogn. Sci.* **8**, 418–425 (2004).
11. Newman, M. E. J. *SIAM Rev.* **45**, 167–256 (2003).
12. Porter, M. A., Onnela, J. P. & Mucha, P. J. *Not. Am. Math. Soc.* **56**, 1082–1097 (2009); go.nature.com/2jg9dgg
13. Fortunato, S. *Phys. Rep.* **486**, 75–174 (2010).

This article was published online on 19 June 2018.

particular. But much less is known about how GPCRs selectively activate inhibitory Gα proteins, which include G<sub>i1</sub>, G<sub>i2</sub>, G<sub>i3</sub> and G<sub>o</sub>, and are collectively known as G<sub>i/o</sub>.

The four papers in this issue report structures of G<sub>i/o</sub>-bound GPCRs obtained using cryo-electron microscopy: Koehl *et al.*<sup>2</sup> (page 547) report the structure of the μ-opioid receptor bound to G<sub>i1</sub>; Draper-Joyce *et al.*<sup>3</sup> (page 559) describe the adenosine A<sub>1</sub> receptor in complex with G<sub>i2</sub>; García-Nafria *et al.*<sup>4</sup> (page 620) report the 5HT<sub>1B</sub> receptor bound to G<sub>o</sub>; and Kang *et al.*<sup>5</sup> (page 553) reveal the structure of the light receptor rhodopsin in complex with G<sub>i1</sub>. The G-protein activation cycle involves the binding and release of nucleotides to and from the G proteins, and all of the reported structures capture the receptors bound to the nucleotide-free state of their respective G proteins.

In some respects, the four structures are similar to those of the previously published GPCR–G<sub>s</sub> complexes<sup>7,8</sup>, probably because G<sub>s</sub>- and G<sub>i/o</sub>-containing complexes have the same overall conformation at the stage of the G-protein activation cycle captured by the structures. Nevertheless, the G<sub>i/o</sub>-containing structures reveal striking differences at the receptor–G-protein interface when compared with the G<sub>s</sub>-containing structures. For example, there are no interactions between the receptors and the Gβ subunits in the G<sub>i/o</sub>-containing structures.

The four structures uncover several key interactions at the GPCR–G<sub>i/o</sub> interface mediated by the α5 helix — an α-helix structure in the carboxy terminus of Gα subunits. It is