

New techniques are coming into play to halt malevolent proteins in their tracks.

THE BIOTHREAT HUNTERS

A cybersecurity-inspired exercise reveals a zero-day vulnerability. **By Matthew Hutson**

DNA-synthesis firms routinely use biosecurity-screening software to ensure that they don't inadvertently create dangerous sequences. But a paper published in *Science* on 2 October describes a potential vulnerability in this workflow¹.

It details how protein-design strategies aided by artificial intelligence (AI) could circumvent the screening software that many DNA-synthesis firms use to ensure that they avoid unintentionally producing sequences encoding harmful proteins or pathogens.

The researchers used an approach from the cybersecurity world: 'red teaming', in which one team attempts to break through another's defences (with their knowledge). They found that some screening tools were unprepared to catch AI-generated protein sequences that recreate the structure, but not the sequence, of known biotreats, says Eric Horvitz, chief

scientific officer at Microsoft in Redmond, Washington. This is a type of zero-day vulnerability – one that, in the cybersecurity world, blindsides software developers and users. "The diversified proteins essentially flew through the screening techniques" that were tested, Horvitz says.

After the developers patched their software to address the new threat, the tools performed much better, flagging all but about 3% of malicious sequences in a larger second attempt.

The impetus for the study is researchers' rapidly growing ability to create new, custom proteins. Armed with AI-powered tools such as RFdiffusion and ProteinMPNN, researchers can now invent proteins to attack tumours, defend against viruses and break down pollutants. David Baker, a biochemist at the University of Washington in Seattle, whose team developed both RFdiffusion and ProteinMPNN, won a share of the 2024 Nobel Prize in Chemistry for

his pioneering work in this area.

But biodesign tools could have other uses – not all of them noble. Someone might intentionally or accidentally create a toxic compound or pathogen, putting many people at risk. The Microsoft-led project aims to prevent that possibility, focusing on a key checkpoint: synthesizing the DNA strands that encode these proteins. Researchers identified gaps in the screening of risky sequences and helped DNA-synthesis providers to close them. But as AI for protein design advances, defences, too, must evolve.

Moment of panic

Horvitz has long recognized that AI, like all technologies, has both good and bad applications. In 2023, motivated by concerns about potential misuse of AI-based protein design, he, Baker and others organized a workshop at the University of Washington to hammer out responsible practices. Horvitz asked Bruce Wittmann, an applied scientist at Microsoft, to create a concrete example of the threat.

Proteins, built of amino acids, are the workhorses of the cell. They are first written in the language of DNA – a string of nucleotides, denoted by A, C, G and T, whose order defines the sequence of amino acids. To create a protein, researchers specify the underlying nucleotide sequence, which they send to a DNA-synthesis company. The provider uses biosecurity screening software (BSS) to look for similarities between the new sequence and known sequences of concern – genes that encode, say, a toxin. If nothing is flagged, the provider creates the requested DNA and mails it back.

Horvitz and Wittmann wanted to see how porous such screening was. So, Wittmann adapted open-source AI protein-design software to alter the amino-acid sequence of a protein of concern while retaining its folded, 3D shape – and, potentially, its function. It's the protein-design equivalent of paraphrasing a sentence, Wittmann says. The AI designed thousands of variants. Horvitz and Wittmann then reached out to two synthesis providers and asked them to use their BSS tools to test the sequences. One was Twist Bioscience in San Francisco, California, which used ThreatSeq from Battelle in Columbus, Ohio; the other was Integrated DNA Technologies (IDT) in Coralville, Iowa, which uses FAST-NA Scanner from RTX BBN Technologies in Cambridge, Massachusetts. The result: the tools were porous, indeed.

Jacob Beal, a computer scientist at BBN, recalls a "moment of panic" looking at one of the tools: "Oh my goodness, this just goes straight through everything, like butter."

Because the findings could have been dangerous in the wrong hands, the team began by sharing them with a small circle of people, including select workshop attendees; US government biosecurity officials; and James Diggans, the chair of the International Gene Synthesis Consortium (IGSC), a coalition of synthesis

Work / Technology & tools

providers, formed in 2009 to create and share standards for screening both sequences and customers.

“The results of the framing study were not a huge surprise,” says Nicole Wheeler, a microbiologist then at the University of Birmingham, UK, and a co-author of the report. But “the study gave a clear indication of the scale of the problem today and data we could use to start testing and improving our screening tools”.

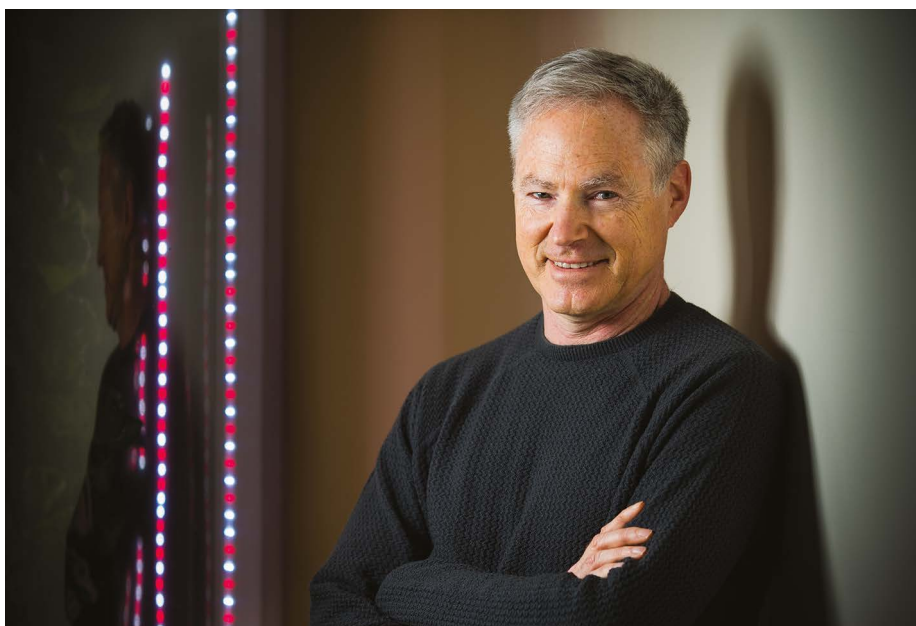
Horvitz and Wittmann then conducted a larger study. They started with 72 proteins of concern – both toxins and viral proteins – and generated tens of thousands of variants of the amino-acid sequences. As before, they ran the design software in two modes, one of which kept amino acids untouched at key locations. This mode increased the chance not only that the proteins would retain the functionality of the unaltered template proteins that they were emulating, but also that they’d be flagged by the BSS. Then, they reverse-translated the amino-acid sequences into DNA, which they sent to four BSS providers who were in on the exercise.

The team also scored the variant proteins for predicted risk. Proteins that exceeded a threshold on two measures were deemed dangerous. First, the proteins needed to be structurally similar (on the basis of computer simulations) to the template proteins. Second, the software needed to have high confidence in the predicted structure, indicating that the protein was likely to fold up properly and be functional. The researchers never actually made the toxic proteins, but in work posted to the preprint server bioRxiv in May², they synthesized some benign ones generated through their design method. They found that their metrics accurately predicted when a protein variant would maintain functionality, suggesting that at least some of the dangerous protein variants would have been functional, too. (But perhaps not many; most of the synthesized variants of benign proteins were inactive.)

Overall, of the proteins that Horvitz and Wittmann deemed most dangerous, the patched BSS caught 97%, while keeping the false-positive rate under 2.5%.

Diggans, who is also the head of biosecurity at Twist, says that the BSS tools that they use were patched in different ways during the *Science* study. In one case, developers used Wittmann’s sequences to fine-tune a machine-learning model; in others, they lowered the statistical-significance threshold for similarity to cast “a wider net”, now that they knew the degree to which AI could change sequences.

Beal, at BBN, says that FAST-NA Scanner works differently. Before the red-teaming exercise, it looked for exact matches between short substrings of nucleotides and the sequences of genes encoding proteins of concern. After being patched, it scans for exact matches only at locations known to be important to a protein’s functionality, allowing for



Eric Horvitz, chief scientific officer at Microsoft.

DAN DELONG

harmless variation elsewhere. The company uses machine learning to generate diverse new sequences of concern, then identifies the important parts of their structures on the basis of similarities between those sequences. Some of the providers have since made further patches on the basis of this work.

Redacted detail

Horvitz and Wittmann teamed up with co-authors, including Wheeler, Diggans and Beal, to write up and share the results. Some colleagues felt the authors should provide every detail, whereas others said they should share nothing. “Our first reaction was, ‘Anybody in the field would know how to do this kind of thing, wouldn’t they?’” Horvitz says. “And even senior folks said, ‘Well, that’s not exactly true.’ And so that went back and forth.”

In the end, they posted a version of their white paper on the preprint server bioRxiv in December, with key details removed. It doesn’t describe the proteins they modified (the *Science* version of the paper lists them), the design tools they used or how they used them. It also omits a section on common BSS failures and glosses over obfuscation techniques – ways to design sequences that won’t raise flags but that produce DNA strands that can easily be modified after synthesis to become more dangerous.

For the published version, the authors worked with journal editors to create a tiered system for data access. Parties must apply through the International Biosecurity and Biosafety Initiative for Science (IBBIS) in Geneva. (The highest-risk data tier includes the study’s code.) “I’m really excited about this,” Tessa Alexanian, a technical lead at IBBIS, said in a press briefing on 30 September. “This managed-access programme is an experiment, and we’re very eager to evolve our approach.”

“There are two communities, which each have very well-grounded principles that both apply here and are in opposition to one another,” Beal says. In the cybersecurity world, people often share vulnerabilities, so they can be patched widely; in biosecurity, threats are potentially deadly and difficult to counter, so people prefer to keep them under wraps. “Now we’re in a place where these two worlds overlap.”

Regulation

Even if screening tools work perfectly, bad actors could still design and build dangerous proteins. There are no laws requiring DNA-synthesis providers to screen orders, for instance. “That’s a scary situation,” says Jaime Yassif, who runs the global biosecurity programme at the Nuclear Threat Initiative (NTI), a non-profit organization in Washington DC. “Not only is screening not required, but the cost of DNA synthesis has been plummeting exponentially for years, and the cost of biosecurity has been basically fixed, so the profit margins on DNA synthesis are pretty thin.” To maximize profit, companies could skimp on screening.

In 2020, the NTI and the World Economic Forum organized a working group to make DNA-synthesis screening more accessible to synthesis firms. The NTI began building a BSS tool called the Common Mechanism, and last year it spun off of IBBIS, which now manages the tool. (Wheeler was the technical lead who developed it.) The Common Mechanism is free, open-source software that includes a database of concerning sequences and an algorithm that detects similarities between those sequences and submitted ones. Users can integrate more databases and analysis modules as they become available.

Still, some scientists think that regulations are necessary. In 2010, the US Department of

Health and Human Services issued guidelines recommending that providers of synthetic double-stranded DNAs screen both sequences and customers, but screening was voluntary. In 2023, former US president Joe Biden issued an executive order on AI safety that, among other things, required researchers who receive federal funding and order synthetic DNA to get it only from providers that screen the orders.

The aim wasn't to stop federally funded researchers from becoming terrorists, Yassif says; it was to add another intervention point to safeguard well-intentioned research that might result in a lab leak or lead to published work that informs terrorists. In any case, President Donald Trump rescinded the order when he took office in January. (An executive order issued on 5 May that halts 'gain of function' pathogen research, also directs the Director of the Office of Science and Technology Policy to "revise or replace" the 2024 Framework for Nucleic Acid Synthesis Screening – a product of the 2023 executive order – to ensure that it "effectively encourages providers of synthetic nucleic acid sequences to implement comprehensive, scalable, and verifiable synthetic nucleic acid procurement screening mechanisms to minimize the risk of misuse".)

Beyond DNA sequences themselves, Yassif says that regulators should look at protecting protein-design software and other biological AI models against misuse. "It's so important to get this right, and DNA-synthesis screening can't be the single point of failure." In 2023, the NTI released a report on AI in the life sciences, based on interviews with 30 specialists. It floated several ideas, including having protein-design AI models screen user requests, restricting the training data, requiring the evaluation of model

safety and controlling model access. A correspondence in *Nature Biotechnology* earlier this year recommended similar safeguards³.

But regulations to protect biological AI models against misuse could be difficult to iron out, Yassif says, because people disagree on risks and rewards. Participants at the University of Washington workshop had a hard time agreeing on a community statement, notes Ian Haydon, the head of AI policy at the university's Institute for Protein Design (which Baker directs). "It's a document that's signed by scores of professors who famously can be a bit stubborn," Haydon says. "It's a bit like herding cats." As a result, its commitments are vague. The biggest area of contention, Haydon says, involved open-source software. "We had people unwilling to sign the language that we arrived at for opposing reasons," he says: some thought it was too supportive of openness, and others thought it was not supportive enough.

The risks of sharing design tools are obvious. Sharing screening tools is also risky, because

"The challenge is: how do we deal with the extra threats in a way that doesn't create an info hazard?"

people who want to synthesize dangerous sequences might work out where the blind spots are and potentially exploit them. The databases in IBBIS's Common Mechanism include well-known proteins of concern, but not some of the more obscure ones. One idea is to send a list of those proteins to approved recipients, but "invariably things will leak", Yassif says. "The challenge this community is facing is: how do we deal with the extra threats beyond the baseline that's publicly known in a way that doesn't create an info hazard?" she says. "That's an unsolved problem."

'Bit of an art'

Even if all synthesis providers did screening, there's a potential workaround: would-be bio-terrorists can buy a synthesis device, although benchtop versions are error-prone and make relatively short segments of DNA (called oligonucleotides) that need to be pieced together. "Oligo synthesis is a bit of an art," Diggans says.

But state-of-the-art technology is changing rapidly. In 2023, the NTI issued a report warning that benchtop synthesizers might be able to build complete viral genomes in as little as a decade. The report recommended regulation. One idea is to require benchtop machines to implement screening internally or over the cloud. But "if there's hardware and software, it can be hacked", says James Demmitt, chief executive of Biolytic Lab Performance, a biotech company in Fremont, California, that makes DNA-synthesis hardware.

That said, defences don't need to be perfect to be effective. "I'm not aware of any solution that is 100% bulletproof," Yassif says. The aim is to "make it harder to exploit this technology to cause harm, shrink the number of people that have the capacity to actually exploit this and do something really dangerous, increase the odds of getting caught, increase the odds of failure. That's what success looks like." According to Demmitt, "biosecurity screening does a good job stopping accidental or casual misuse. By forcing folks to go through bigger, pricier hoops, it prevents many would-be dabblers from drifting into dangerous territory."

And there are more technical hurdles facing bad actors. Rarely is DNA itself a danger; people need to engineer sequences into cells or viruses to manufacture toxins or produce self-replicating pathogens. That requires biological know-how and equipment beyond many people's means. Even for specialists, there's a huge gap between designing a protein or virus and knowing its effect in people. That's why pandemic prediction is so difficult. Scientists find viruses in the wild that seem dangerous, but few infect people, fewer spread between them and even fewer make them sick.

Whatever the chances of someone designing something deadly, specialists say we should remain vigilant, just as in cybersecurity – but the cat-and-mouse games are different in one regard. "In the cyber world, you have a lot of people looking to exploit these systems," Diggans says, "from the 'script kiddie' teenagers looking to do it for fun, all the way to the multinational crime syndicates." He continues: "It is vanishingly rare to have anyone who wants to exploit biotechnology for nefarious purposes. That is both good – because we don't want people exploiting biotech – but it is also hard, because it gives us very few signals against which to build defences." In March, the US National Academies of Sciences, Engineering, and Medicine recommended that more research into methodologies for nucleic-acid-synthesis screening is needed.

As that happens, the field can continue to take lessons from cybersecurity specialists, who have been going toe-to-toe with bad actors for decades. "What stands out to me" in the new paper, Haydon says, "is the way they wove in practices and precedents from cybersecurity", such as letting providers build patches before publicizing their findings. As the field develops, providers will need to keep upping their game. As a "Microsoft person", Horvitz is reminded of the Windows update model. "This will never be ending," he says.

Matthew Hutson is a freelance science writer in New York City.



Applied scientist Bruce Wittmann.

1. Wittmann, B. J. et al. *Science* **390**, 82–87 (2025).
2. Ikononova, S. P. et al. Preprint at bioRxiv <https://doi.org/10.1101/2025.05.15.654077> (2025).
3. Wang, M. et al. *Nature Biotechnol.* **43**, 845–847 (2025).